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Isochore chromosome maps of the human genome

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Abstract

The human genome is a mosaic of isochores, which are long DNA segments ($\gg$ 300 kbp) relatively homogeneous in G + C. Human isochores were first identified by density-gradient ultracentrifugation of bulk DNA, and differ in important features, e.g. genes are found predominantly in the GC-richest isochores. Here, we use a reliable segmentation method to partition the longest contigs in the human genome draft sequence into long homogeneous genome regions (LHGRs), thereby revealing the isochore structure of the human genome. The advantages of the isochore maps presented here are: (1) sequence heterogeneities at different scales are shown in the same plot; (2) pair-wise compositional differences between adjacent regions are all statistically significant; (3) isochore boundaries are accurately defined to single base pair resolution; and (4) both gradual and abrupt isochore boundaries are simultaneously revealed. Taking advantage of the wide sample of genome sequence analyzed, we investigate the correspondence between LHGRs and true human isochores revealed through DNA centrifugation. LHGRs show many of the typical isochore features, mainly size distribution, G + C range, and proportions of the isochore classes. The relative density of genes, Alu and long interspersed nuclear element repeats and the different types of single nucleotide polymorphisms on LHGRs also coincide with expectations in true isochores. Potential applications of isochore maps range from the improvement of gene-finding algorithms to the prediction of linkage disequilibrium levels in association studies between marker genes and complex traits. The coordinates for the LHGRs identified in all the contigs longer than 2 Mb in the human genome sequence are available at the online resource on isochore mapping: http://bioinfo2.ugr.es/isochores.

Keywords: Isochore maps; Compositional segmentation; Chromosome domains; Comparative genomics; Alus; Long interspersed nuclear elements; Single nucleotide polymorphisms

1. Introduction

The availability of the human genome draft sequence offers an unprecedented opportunity to bring sequence patterns into line with the chromosome structures revealed by modern molecular cytogenetics, such as chromosome domains or high-resolution chromosome bands. Isochores – long DNA segments ($\gg$ 300 kbp) fairly homogeneous in G + C, revealed by analytical ultracentrifugation of bulk DNA (Macaya et al., 1976; Bernardi et al., 1985; Bernardi, 1995, 2000) – may be the structures linking both organization levels. In fact, isochores have been successfully related to chromosome bands (Saccone et al., 1993).

One conventional way to visualize sequence heterogeneity is the moving-window approach. This simple technique consists of sliding a window of arbitrary length along the sequence, and then computing the GC content of each window. This procedure dates from the earliest times of sequence analysis when only short, and often homogeneous, sequences were available. However, with the discovery that eukaryotic genomes are multi-scale complex systems made up of fairly homogeneous isochores of different composition (Macaya et al., 1976; Bernardi et al., 1985; Bernardi, 2000) and with the subsequent finding of long-range correlations in eukaryotic DNA sequences (Li and Kaneko, 1992; Peng et al., 1992; Voss, 1992; Bernaola-Galván et al., 2002a), this

Abbreviations: LHGR, long homogeneous genome region; bp, base pair; kbp, kilobase pair; G + C, guanine plus cytosine content; SNP, single nucleotide polymorphism; MY, millions of years; SINE, short interspersed nuclear element; LINE, long interspersed nuclear element.

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practice becomes untenable. Sliding a window of arbitrary length and step over long, heterogeneous and correlated sequences may lead to misleading results (see Li, 2001, for a recent review). However, GC-plots routinely accompany the publication of every new genome sequence, the long-range patterns being identified only by eye. This happens, for example, with the 'isochores' tentatively identified on human chromosomes 21 (Hattori et al., 2000) and 22 (Dunham et al., 1999).

Two other more recent techniques (Nekrutenko and Li, 2000; Häring and Kypr, 2001), also based on moving windows, use the random (uncorrelated) model to test sequence homogeneity. The pitfalls in such an approach have already been noted (Bernardi, 2001; see also Li et al., 2002). A key problem is that moving windows do not enable the accurate location of isochore boundaries before carrying out the homogeneity test. Therefore, it is not surprising that one of these techniques (Nekrutenko and Li, 2000) failed to detect the only isochore boundary experimentally characterized to date (Fukagawa et al., 1995, 1996; Stephens et al., 1999), while the other (Häring and Kypr, 2001) was unable to detect any isochores in the human chromosomes 21 and 22.

An alternative tool to analyze genome heterogeneity is compositional segmentation (Bernaola-Galván et al., 1996, 2001; Li et al., 1998; Román-Roldán et al., 1998; Oliver et al., 1999, 2001; Li, 2001). Domains of all sizes can be simultaneously detected by this method, and isochore boundaries can be accurately determined to single base pair resolution.

A recently derived hierarchical segmentation method (Oliver et al., 2001; Román-Roldán et al., 2002) is used here to divide the longest contig of each human chromosome into non-overlapping, relatively homogeneous genome regions, called long homogeneous genome regions (LHGRs). To investigate to what extent these regions may correspond to the true isochores identified by the Bernardi group through DNA centrifugation, we analyze here several LHGR features, such as size distribution, G + C range, and proportions of the different compositional classes in a wide sample of human genome sequence. We also analyzed the relative densities of genes, Alu and long interspersed nuclear element (LINE) repeats and the different types of single nucleotide polymorphisms (SNPs) in these regions.

2. Materials and methods

Different freezes, from October 2001 to February 2002, of the public human genome draft sequence available at NCBI (Lander et al., 2001; ftp://ncbi.nlm.nih.gov/genomes/H_sapiens) were used to compile information for different parts of this work. All the contigs longer than 2 Mb in the human genome were segmented using our hierarchical algorithm (for a complete list see the online resource on isochore mapping: http://bioinfo2.ugr.es/isochores). The

Table 1
Longest human contigs by chromosome analyzed in this study (NCBI October 2001 freeze[a])

Chromosome	Accession	Contig version	Contig length (bp)
1	NT_004424	6	6,311,978
2	NT_005375	6	4,746,219
3	NT_005927	6	19,259,936
4	NT_006204	6	5,458,445
5	NT_006907	6	4,272,479
6	NT_007592	6	19,443,354
7	NT_007819	6	12,615,535
8	NT_008271	6	3,868,249
9	NT_008413	6	8,724,786
10	NT_008609	6	8,702,417
11	NT_009151	6	24,188,643
12	NT_009714	6	5,170,685
13	NT_024524	6	10,245,455
14	NT_025892	5	16,139,217
15	NT_010194	6	10,898,583
16	NT_010604	6	4,049,516
17	NT_010718	6	8,843,538
18	NT_010895	6	4,073,989
19	NT_026483	4	4,069,655
20	NT_011362	6	26,179,448
21	NT_011512	4	28,511,026
22	NT_011520	8	23,083,944
X	NT_011687	6	6,615,739
Y	NT_011875	7	9,946,786
			Total: 275,419,622 (8.6% of the genome)

A complete list of the contigs analyzed can be found at the online resource on isochore mapping: http://bioinfo2.ugr.es/isochores.

[a] ftp://ncbi.nlm.nih.gov/genomes/H_sapiens.

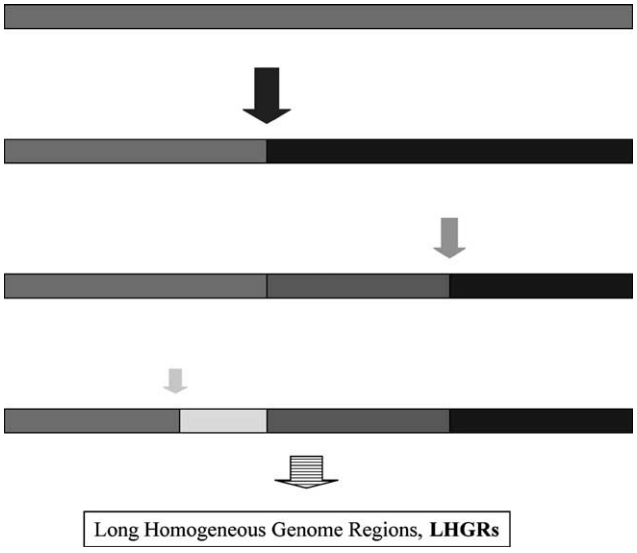

Fig. 1. Schematic representation of the segmentation algorithm used to locate LHGRs on sequence contigs. The successive cuts are given in a hierarchical way: at each scale, the cut maximizing the overall compositional complexity of the sequence is chosen, a procedure equivalent to maximizing the statistical significance of each cut (see Oliver et al., 2001, for details).

LHGRs identified on the longest contigs of each chromosome (Table 1) were used for most of the statistical comparisons described in this paper.

The segmentation algorithm used here (Oliver et al., 2001, 2002; Román-Roldán et al., 2002) is based on the original method developed by our group (Bernaola-Galván et al., 1996, 1999, 2000; Román-Roldán et al., 1998; Oliver et al., 1999; Grosse et al., 2002), but with several improvements (all aimed at addressing the specific isochore mapping problem). A schematic representation of the improved method is shown in Fig. 1. Two features should be emphasized:

(a) The cuts on the sequence are made one by one, in a hierarchical way, which may be more appropriate in searching for homogeneous segments in the long-range correlated, fractal landscape of eukaryotic DNA. In such a multi-scale landscape, the statistical significance of isochore boundaries may depend on the scale being considered. The hierarchical procedure guarantees the choice of the most significant cut at each scale.
(b) Short-scale sequence heterogeneity below 3 kbp is filtered out; this coarse graining of the sequence is a requirement imposed by the experimental characterization of isochores through DNA centrifugation (Bettecken et al., 1992; Bernardi, 2000). Filtering out heterogeneities below 3 kbp is also justified by the analysis of correlations in human chromosomes 20 and 21 (Bernaola-Galván et al., 2002a), which show a clear shift between two heterogeneity regimes just at this scale.

Therefore, our procedure tries to identify isochores by closely following Bernardi's early definition, i.e. 'fairly homogeneous regions', which implies accepting a certain level of internal heterogeneity. The 'strict isochores', unsuccessfully searched for by other authors (Lander et al., 2001; Häring and Kypr, 2001), simply cannot exist in natural DNA (Bernardi, 2001).

Parameter settings were as in the previous work (Oliver et al., 2001), i.e. a tract length of 3 kbp was used for coarse graining of GC content, and a 0.05 threshold was set for the P value in t-tests. These settings provide a high stability in detecting isochores in the human MHC region: the same isochore structure was obtained with coarse graining ranging from 2 to 30 kbp (see Fig. 2 in Oliver et al., 2001).

This segmentation method has been used to accurately predict the boundary between classes II and III of the human MHC region (Oliver et al., 2001), the only isochore boundary experimentally determined to date (Fukagawa et al., 1995; Stephens et al., 1999; The MHC Sequencing Consortium, 1999). The method has also been used to uncover isochore-like regions in other eukaryotic genomes (Oliver et al., 2001). More recently, we are also using this method to explore sequence heterogeneity in prokaryotic genomes (Bernaola-Galván et al., 2002b).

Gene ('CDS' line) and SNP ('variation' line) coordinates were taken from contig annotations. Chromosome contigs were scanned for Alu and LINE repeats using the program RepeatMasker (http://repeatmasker.genome.washington.edu), which identifies full-length and partial members of all the known repeat families represented in RepBase (Jurka, 2000; http://www.girinst.org/~server/repbase.html).

3. Results and discussion

3.1. Isochore chromosome maps

We applied our segmentation algorithm to the longest contig of each human chromosome available at the NCBI web server. As an example, the isochore chromosome maps for the longest contigs of chromosomes 21 and 22 are shown in Figs. 2 and 3. A more compact representation is used in Fig. 4 to show the isochore maps of the 24 longest contigs in the human chromosome complement. Isochore chromosome maps of every long human contig are regularly updated at the online resource on isochore mapping: http://bioinfo2.ugr.es/isochores. These maps graphically display the mosaic organization of the human genome (Bernardi et al., 1985; Bernardi, 2001; Pavlícek et al., 2001), composed by many regions of fairly homogeneous GC contents (see Li et al., 2002 for a recent reassessment of isochore homogeneity). The advantages of these maps over previous approaches based on moving windows are: (1) heterogeneities at very different scales are shown in the same plot; (2) pair-wise differences in GC content between adjacent regions are all statistically significant; (3) the

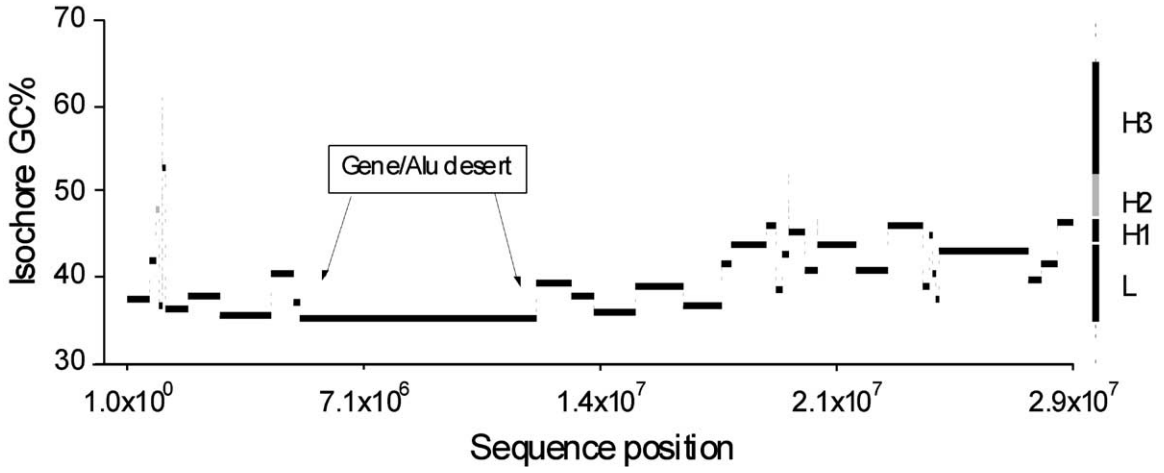

Fig. 2. Isochore chromosome map of the longest contig of human chromosome 21. The gene and Alu desert of 7.1 Mb is indicated by the arrows.

isochore boundaries are accurately defined to single base pair resolution; and (4) both gradual and abrupt isochore boundaries can be simultaneously revealed. As recently stressed by Bernardi (2001), this generalized mosaic structure along all the human chromosomes contradicts the suggestion (Eyre-Walker and Hurst, 2001) that the isochore structure accounts for 'only some parts' of the genome.

3.2. The relative amounts of DNA in the different compositional families

The LHGRs we found were classified into compositional families on the basis of their respective GC content, according to the GC values of Zoubak et al. (1996). The relative amounts of DNA in L, H1, H2 and H3 LHGRs in the longest contig of each human chromosome (Fig. 5) were fairly similar to the proportions experimentally found in the entire human genome by DNA centrifugation (e.g. Zoubak et al., 1996).

3.3. Statistical features of LHGRs

The size distribution of LHGRs, the distribution of GC contents and the GC differences between adjacent LHGRs are shown in Fig. 6. The LHGR size distribution was strongly skewed, with the highest value being 7.1 Mb (corresponding to the gene desert of chromosome 21) and an average size of 463 kbp. Many of the smaller LHGRs may correspond to GC-skewed repeats (Alus, LINEs), CpG islands or attached scaffold regions. LHGR GC content ranges from 30.8% to 64.3%, thus being consistent with the known GC range in isochores (Bernardi, 2001). The GC differences between adjacent isochores range from 1.5% to 24.5%. The smaller compositional shifts were noted between L-L adjacent LHGRs, while the greater ones occurred between L-H LHGRs. We observed, therefore, both gradual and abrupt GC shifts between adjacent isochores, depending on the neighbors considered. This observation contrasts with all previous reports in which only abrupt isochore boundaries were found (see, for example,

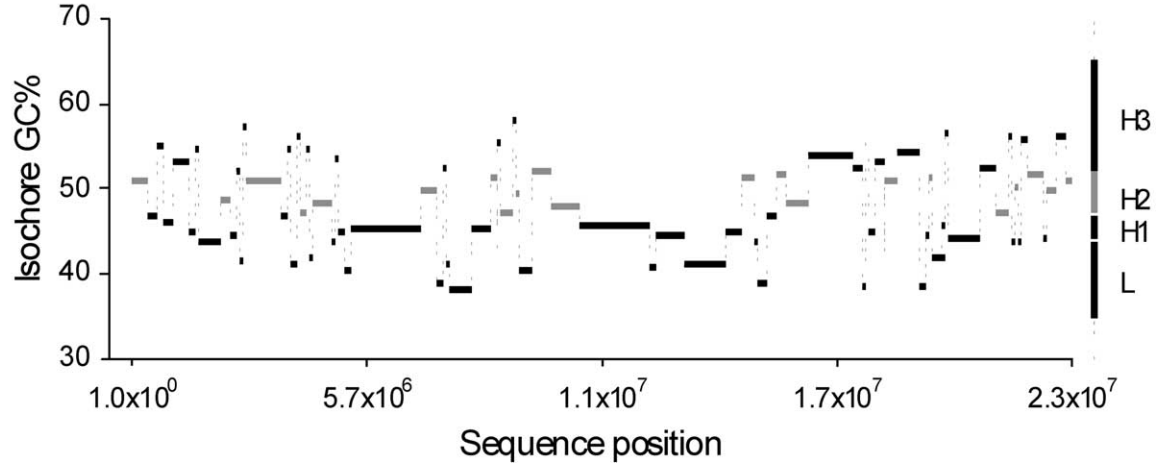

Fig. 3. Isochore chromosome map of the longest contig of human chromosome 22.

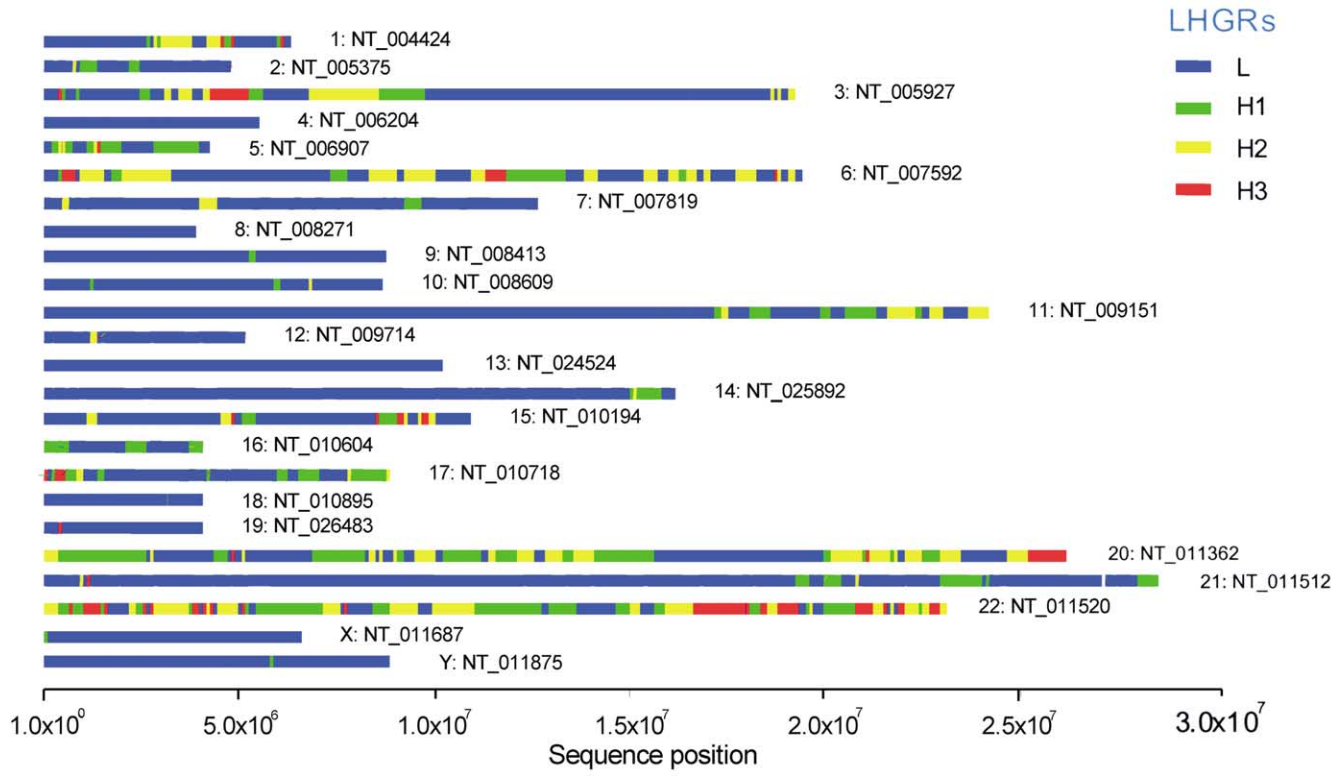

Fig. 4. Isochore chromosome maps of the longest contig in the human chromosome complement. The October 2001 freeze of NCBI contigs was used.

Fukagawa et al., 1995; Stephens et al., 1999). Note that the moving-window plot used by most authors only allows for the detection of abrupt transitions, while our segmentation method can reveal both gradual and abrupt isochore boundaries.

3.4. LHGR size variation with GC content

The different LHGR families show a strong variation in size, depending on the GC content, GC-poor LHGRs being significantly larger than GC-rich ones (Table 2). This

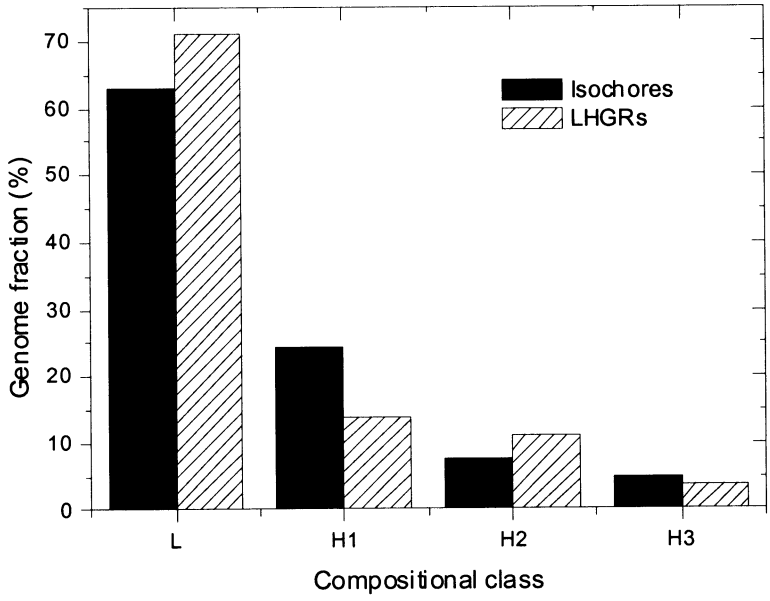

Fig. 5. The relative amounts of DNA in the different compositional LHGR families. The LHGRs in the longest contig of each chromosome (NCBI, October 2001 freeze), amounting to a total of 275.4 Mb (8.6% of the genome), were compared to the isochores detected by DNA centrifugation in the entire genome (Zoubak et al., 1996). LHGR G + C ranges (taken from Zoubak's paper) were: L1-L2 (GC% $<$ 44), H1 (44 $\leq$ GC% $<$ 47), H2 (47 $\leq$ GC% $<$ 52) and H3 (GC% $\geq$ 52).

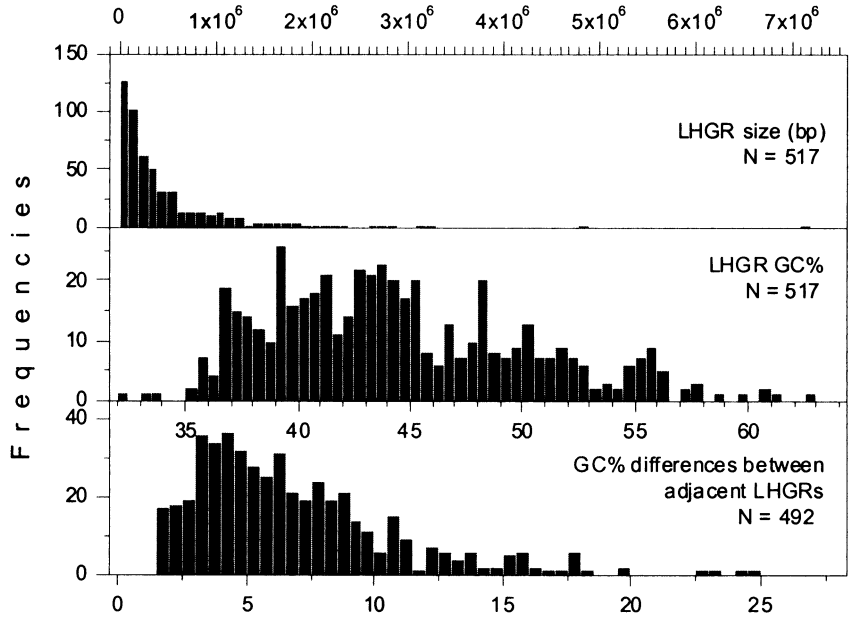

Fig. 6. Size distribution (above), GC content (middle) and GC differences between adjacent LHGRs in the longest contig of each human chromosome. A total of 517 LHGRs were considered. Contigs were taken from the NCBI October 2001 freeze, amounting to a total of 275 Mb (8.6% of the genome).

relationship was previously noted for the isochores detected by DNA centrifugation (Bettecken et al., 1992; Pilia et al., 1993; De Sario et al., 1996, 1997).

3.5. Variation of gene density in human LHGRs

In isochores detected by DNA centrifugation, Bernardi and coworkers (Bernardi et al., 1985; Mouchiroud et al., 1991; Zoubak et al., 1996; Bernardi, 2000) observed that gene density increases from a very low average in L isochores to a 20-fold higher level in H3 isochores. The recent release of the human genome draft sequence (Lander et al., 2001; Venter et al., 2001) propitiated a reexamination of this relation; while the first of the analyses, using 20 kbp windows along the assembled sequence, confirms the original observation, the second one, using 50 kbp windows, questioned the relative strength of the correlation. Thus, Venter et al. (2001) found that the correlation between GC content and gene density was not as skewed as observed by Bernardi's group, a higher proportion of genes being located in the GC-poor regions than had been previously observed in isochores. We therefore check this relation by using the human isochore boundaries accurately determined through our segmentation algorithm. Fig. 7 illustrates the close relationship we found between LHGR G + C and gene density (number of genes per kilobase). These results were remarkably similar to those of Bernardi's group (Mouchiroud et al., 1991; Zoubak et al., 1996; Bernardi, 2000, 2001), with our gene density values also falling on two straight lines crossing each other at about 46% GC. The less skewed distribution observed by Venter et al. (2001) may be due to (1) the specific values chosen for the window length and/or step, or (2) a wrong definition of the GC ranges assigned to each isochore family.

Table 2
Sizes of LHGRs (in kb) belonging to different families

LHGR	*N*	Mean	SE	Minimum	Maximum
L	276	615	49	9	7105
H1	84	399	49	16	2293
H2	97	281	29	3	1794
H3	60	144	28	6	1121

An analysis of the variance shows that size differences were statistically significant ($P < 10^{-6}$). The NCBI October 2001 freeze of contigs was used to compile this table.

3.6. Variation in the densities of Alu and LINE repeats

The density of Alu and LINE repeats is known to vary with isochore GC content (Soriano et al., 1983; Smit, 1999; Lander et al., 2001). To investigate if this relation is also true for LHGRs, we analyzed in detail the variations in Alu density along the LHGRs detected by our segmentation algorithm in 131 contigs longer than 3.5 Mb in the human genome (NCBI February 2002 freeze). We found a relationship between LHGR GC content and Alu density. However, the strength of such a relationship depends on the genetic age of the Alu family considered. Fig. 8 shows the average densities of two Alu families of different ages; the genetic ages of Alu families were taken from Kapitonov and Jurka (1996). While the density of the old Alu S family is strongly dependent on the isochore GC content, no

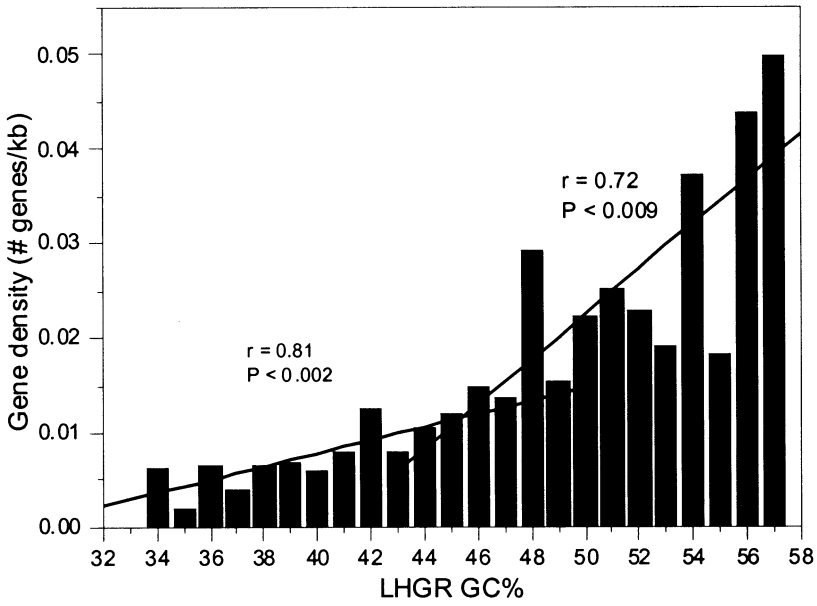

Fig. 7. Gene density vs. LHGR GC content. A total of 1096 genes located on 495 LHGRs from the longest contigs of each human chromosome were used for the comparison.

relationship was found for the youngest Alu Y family. LINE L1 density shows just the opposite pattern to that of the old Alus, being more frequent in L isochores and practically absent in the H3 isochores. Therefore, the density of Alu and LINE repeats in LHGRs follows the patterns previously found for isochores (Soriano et al., 1983; Smit, 1999; Lander et al., 2001).

3.7. Compositional correlations between gene GC content and LHGR G + C

Working with the isochores detected by DNA centrifugation, it has been convincingly shown that the GC content of genes matches the G + C of the isochores harboring them (Bernardi et al., 1985). Figs. 9 and 10 and Table 3 show that

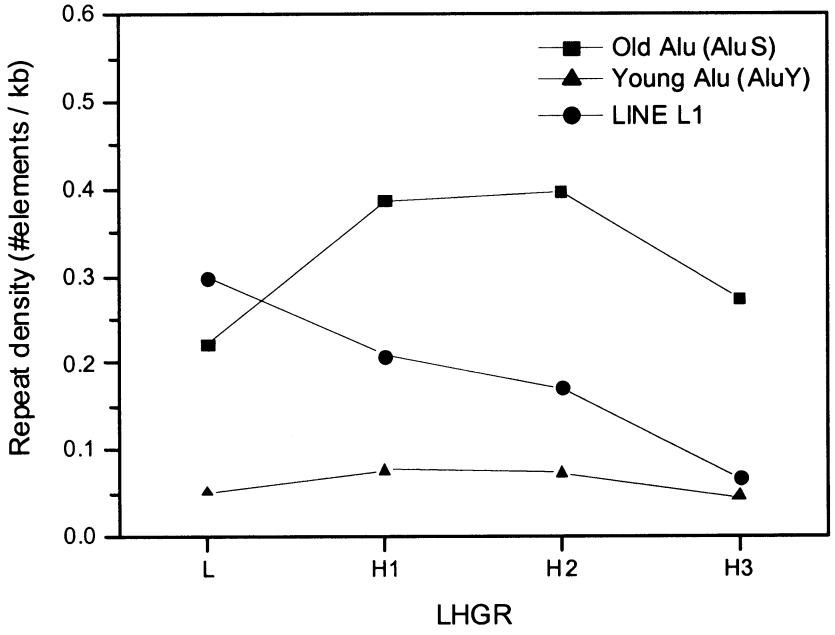

Fig. 8. Alu and LINE densities in the different LHGR families from 131 contigs longer than 3.5 Mb in the human genome (NCBI February 2002 freeze). Around 330,000 Alus and 345,000 LINEs in 2048 LHGRs were used to compile this figure. Old (S) and young (Y) Alus and the older LINE L1 elements were included in the comparison.

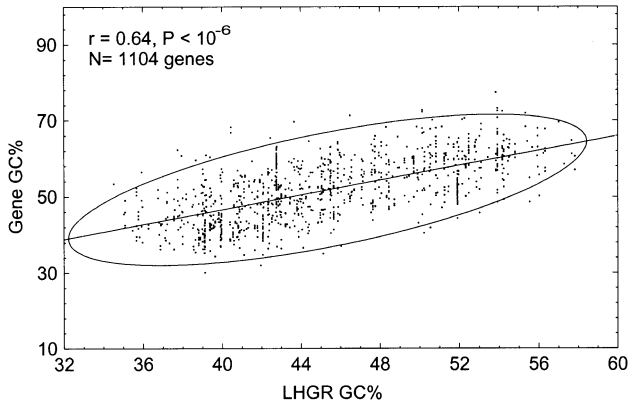

Fig. 9. Gene GC content vs. LHGR G + C ($r = 0.64$, $P < 10^{-6}$). A total of 1104 genes from the longest contig of each chromosome were included in the comparison. The ellipse shows 95% confidence intervals.

this rule also holds for the LHGRs detected by segmenting human contigs, thus being consistent with previous results (see, for example, Eyre-Walker and Hurst, 2001).

3.8. The compositional adjustment of Alus and LINES to LHGR GC content

The study of sequence repeats, such as Alus and LINEs, can also reveal compositional correlations. An advantage of using repeats instead of genes for this purpose is that the genetic ages of the different repeat families are known (Kapitonov and Jurka, 1996; Mighell et al., 1997), and therefore the evolution towards the compositional matching with the host sequence (a process known as 'compositional adjustment'; Oliver et al., 1990; Martínez Zapater et al., 1993) can be analyzed in more detail. Figs. 11–13 show the correlation plots between the GC content of repeats and the G + C of the LHGR harboring them for AluJo (81 MY old), AluSp (37 MY old) and AluYa5 (4 MY old) families, respectively. The strongest correlation, and therefore the best compositional adjustment, was for the oldest AluJo repeats, while no correlation was found for the youngest

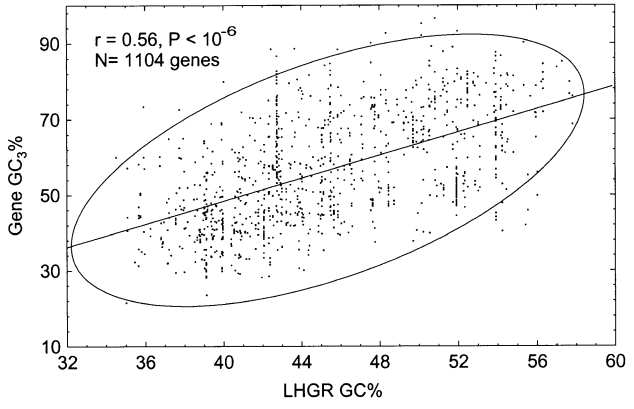

Fig. 10. Gene GC_3 content vs. LHGR G + C ($r = 0.56$, $P < 10^{-6}$). A total of 1104 genes from the longest contig of each chromosome were included in the comparison. The ellipse shows 95% confidence intervals.

Table 3
Gene GC content (%GC) and G + C at third codon positions (%GC_3) in LHGR families

LHGR	*N*	%GC ± SE	%GC_3 ± SE
L	526	47.05 ± 0.29	49.10 ± 0.55
H1	172	52.32 ± 0.48	57.08 ± 0.94
H2	263	55.86 ± 0.36	63.62 ± 0.77
H3	143	60.78 ± 0.45	69.41 ± 0.98

Data from the longest contig of each chromosome.

AluYa5 elements. The trend to increase the compositional adjustment with time is confirmed by the fact that the very old LINE L2 repeats (>120 MY old) show the strongest correlation ($r = 0.73$, Fig. 14). Therefore, the compositional adjustment of Alu and LINE repeats to the isochores harboring them is a time-dependent process, thus fitting well within the framework of the neutral theory of molecular evolution (Kimura, 1983). In this way, Alus and LINEs, taken all together, appear to be neither beneficial nor harmful to the host; adaptive or maladaptive functions, if any, need to be demonstrated on an individual basis.

3.9. SNP density in different LHGR families

The SNP density (SNPs/kilobase) varies considerably among and within human chromosomes, the observed distribution of SNPs in 100 kbp fragments of the draft genome sequence showing far more pronounced variance than expected by chance (Venter et al., 2001).

By using our segmentation algorithm on the longest contig of each human chromosome, and by collecting all the annotated SNPs save those at CpG sites, we also found such a relationship for LHGRs (Fig. 15 and Table 4). A trend for SNP density to increase with LHGR GC level can be appreciated, the differences being significant between L and H3 LHGRs ($P < 10^{-3}$).

Fig. 15 also shows the variation in the densities of the six

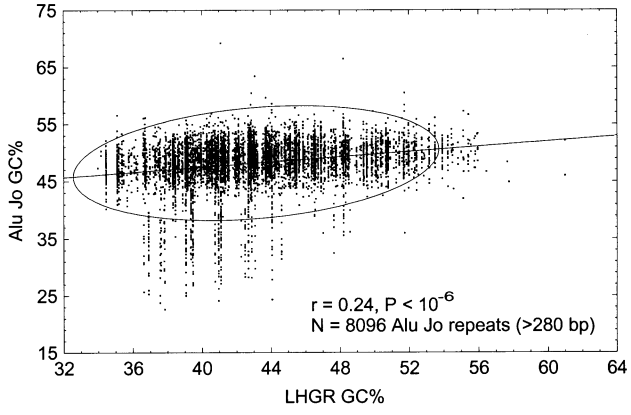

Fig. 11. AluJo GC content vs. LHGR G + C ($r = 0.24$, $P < 10^{-6}$). A total of 8096 AluJo repeats larger than 280 bp located on the longest contig of each chromosome (NCBI October 2001 freeze) were used in the comparison. The ellipse shows 95% confidence intervals.

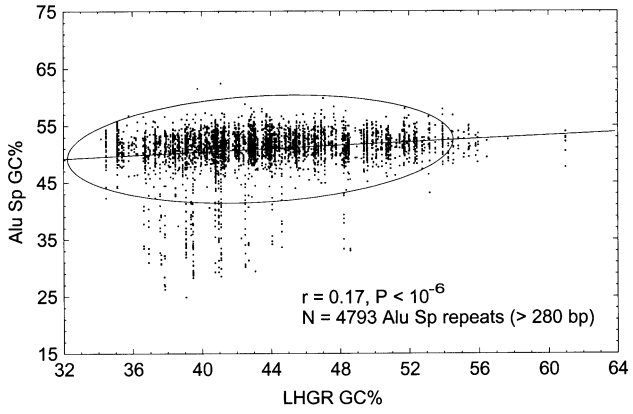

Fig. 12. AluSp GC content vs. LHGR G + C ($r = 0.17$, $P < 10^{-6}$). A total of 4793 AluSp repeats larger than 280 bp located on the longest contig of each chromosome (NCBI October 2001 freeze) were used in the comparison. The ellipse shows 95% confidence intervals.

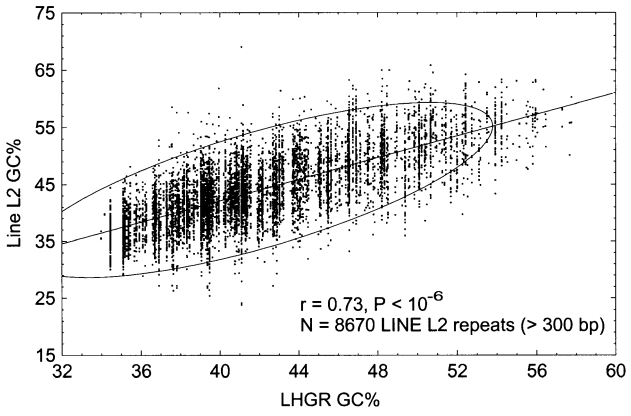

Fig. 14. LINE L2 GC content vs. LHGR G + C ($r = 0.74$, $P < 10^{-6}$). A total of 8670 LINE L2 repeats larger than 300 bp located on the longest contig of each chromosome (NCBI October 2001 freeze) were used in the comparison. The ellipse shows 95% confidence intervals.

possible base changes at the SNPs mapping on the different LHGR families. Considerable variations were observed among the different types of base changes, but, for a given base change, only slight variations among LHGR families were detected.

Lastly, we analyzed the ratio of transition and transversion substitutions in LHGR families (Table 4), and found variations with roughly the typical 2:1 ratio for mammalian genomes (Graur and Li, 2000). A slight trend for the transition/transversion ratio to increase with the LHGR G + C content appeared, although statistical significance was not reached.

3.10. Conclusions

By means of a hierarchical segmentation algorithm, specifically designed to determine the most statistically significant partition of a DNA sequence at each scale, here we have drawn isochore maps defined to single base pair resolution for the longest contig of each human chromosome. The fairly homogeneous regions found (LHGRs) displayed many of the features (G + C range, proportion of isochore classes, size distribution and relationship with gene and Alu densities) of the isochores identified through the centrifugation of vertebrate DNA fragments. The known correlations between different biological features (gene, repeat and SNP densities) and the isochore G + C content were also observed in LHGRs.

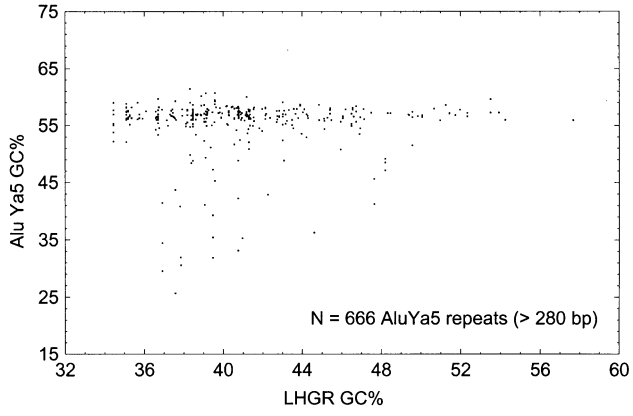

Fig. 13. AluYa5 GC content vs. LHGR G + C. A total of 666 AluYa5 repeats larger than 280 bp located on the longest contig of each chromosome (NCBI October 2001 freeze) were used in the comparison.

The isochore chromosome maps of the human genome presented here show several advantages over previous approaches based on moving windows: (1) sequence heterogeneities at different scales are simultaneously shown; (2) pair-wise differences in GC content between adjacent regions are all statistically significant; (3) isochore boundaries are defined to single base pair resolution; and (4) both gradual and abrupt isochore boundaries are simultaneously revealed.

The computational prescreening of isochore boundaries may have many applications in genomics: (1) the changes in replication timing known to occur at isochore boundaries (Tenzen et al., 1997) can now be exhaustively searched for at the predicted LHGR boundaries; (2) the genomic sequences can now be scanned for gene-rich regions, as we found that gene density depends heavily on the GC content of the LHGRs; (3) improvements in computational gene identification are also expected, as the specific compositional parameters of the corresponding isochores

Table 4
SNP density and ratio of transition to transversion substitutions in LHGR families

LHGR	SNP density (# SNP/kb) ± SE	Transition/transversion rate ± SE
L	0.47 ± 0.02	2.00 ± 0.05
H1	0.51 ± 0.06	2.09 ± 0.08
H2	0.57 ± 0.04	2.17 ± 0.09
H3	0.68 ± 0.07	2.10 ± 0.15

All the annotated SNPs (112,826) in the longest contig of each human chromosome, save those at CpG sites, were analyzed.

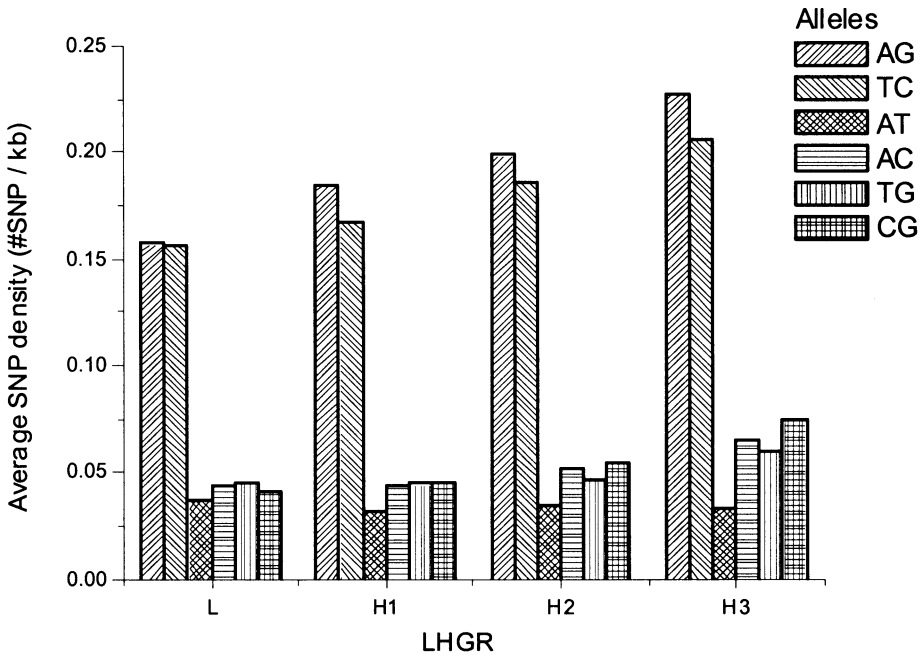

Fig. 15. Densities of different SNPs in LHGR compositional families. All the annotated SNPs in the longest contig of each human chromosome, save those at CpG sites, were analyzed. The densities of the six possible base changes are shown.

can now be taken into account as input for gene-finding programs (Burge and Karlin, 1997); in fact, we have recently shown (Carpena et al., 2002) that the prediction of the coding proportion in a sequence is better when LHGRs, instead of moving windows, are used; (4) in the same way, other programs making use of local compositional parameters to predict sequence patterns, as RepeatMasker (http://repeatmasker.genome.washington.edu), could be improved by considering LHGRs instead of moving windows; (5) the transitions from long-range to short-range linkage disequilibrium can coincide with switches in the isochore pattern (Eisenbarth et al., 2000, 2001); if so, the precise delimitation of isochore boundaries can help to predict the levels of linkage disequilibrium, thereby facilitating association studies, the most powerful current tool for the identification of genes underlying complex traits; and (6) the analysis of isochore chromosome maps in different genomes may allow new insights in the field of comparative genomics.

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